

Mutagenic Properties of Methyl- and Ethylbromophos in Mammals

N. Degraeve,¹ M.-C. Chollet,¹ and J. Moutschen²

¹Laboratoire de Chimie Médicale et de Toxicologie, Institut de Pathologie, B 23, Université de Liège 4000 Sart Tilman par Liège 1, Belgium and ²Laboratoire de Génétique, Université de Liège, 15 rue Forgeur, 4000 Liège, Belgium

Although the mutagenic properties of some organophosphorus pesticides such as dichlorvos and trichlorfon have been extensively investigated (Moutschen et al., 1981), only a few data are available concerning some other substances of the same chemical group. This is the case for methylbromophos (dimethyl 4 bromo 2,5 dichlorophenyl phosphorothionate) and ethylbromophos (diethyl 4 bromo 2,5 dichlorophenyl phosphorothionate), two substances used for insect control on both animals and plants (Brofene, Bromophos, Nexion, Prosol, Nexagan).

The only published data does not reveal any mutagenic activity in Salmonella typhimurium (Lippens et al., 1983), Schizosaccharomyces pombe (Gilot-Delhalle et al., 1983) and Drosophila melanogaster (Benes and Sram, 1969).

The present research investigates the genetic (dominant lethal mutations) and cytogenetic (chromosome damage in bone marrow cells and spermatogonia) effects induced in male mice by an acute treatment with a very high dose (1000 mg/kg) of methylor ethylbromophos. The results are compared to those obtained with the well known mutagen mitomycin C selected as a positive control.

MATERIALS AND METHODS

Methylbromophos and ethylbromophos (Riedel-De Haen, Pestanal[®], purity > 99%) being slightly soluble in water, the solutions were injected as suspensions in peanut oil.

Male mice (Q strain), 3-4 months old, received a single i.p. injection. The doses tested were 1 g/kg for the insecticides and 2 mg/kg for mitomycin C (Sigma Chemical).

Treated males were sacrificed from 12-36 h after the treatment for the analysis of chromosome damage in bone marrow cells and spermatogonia. Microscopic slides were prepared according to the air drying method (Evans et al., 1964).

Table 1: Cytogenetic effects induced in bone marrow cells by methylbromophos (1 g/kg), ethylbromophos (1 g/kg) and mitomycin C (2 mg/kg) (500 metaphases analyzed).

Treatment	Recovery period in h	Percentage of aberrations		
		Fragments	Exchanges	Gaps
Control	-	0.4	0.0	0.4
Mitomycin C	12	7.2	2.6	5.0
	24	45.2	8.2	8.6
	36	7.0	1.6	0.6
Methyl- bromophos	12	0.0	0.0	0.0
	24	0.4	0.0	0.0
	36	0.2	0.0	0.0
Ethyl- bromophos	12	0.2	0.0	0.2
	24	0.4	0.0	0.0
	36	0.6	0.0	0.0

Table 2: Cytogenetic effects induced in spermatogonia by methylbromophos (1 g/kg), ethylbromophos (1 g/kg) and mitomycin C (2 mg/kg).

Treatment	Recovery period in h	Number of metaphases analyzed	Percentage of aberrations		
			Fragments	Exchanges	Gaps
Control	-	386	0.26	0.00	0.26
Mitomycin C	12	132	15.91	9.09	6.82
	24	68	17.65	11.76	4.41
	36	33	27.27	6.06	3.03
Methyl- bromophos	12	143	1.40	0.00	0.70
	24	408	0.74	0.00	0.49
	36	336	0.30	0.00	0.00
Ethyl- bromophos	12	289	1.04	0.00	0.00
	24	366	0.54	0.00	0.27
	36	360	0.28	0.00	0.28

Table 3: Genetic effects induced by methylbromophos (1 g/kg), ethylbromophos (1 g/kg) and mitomycin C (2 mg/kg) in the dominant lethal mutation assay.

Treatment	Recovery period in days	Pregnant females	Corpora lutea per ♀	Implants per ♀	Preimplantation losses per ♀	% CL	Live embryos per ♀	Postimplantation losses per ♀	% IP	Total mortality per ♀	%CL
Control	-	68	9.93	9.15	0.78	7.9	8.78	0.37	4.0	1.15	11.6
Mitomycin C	1-7	11	9.73	9.64	0.09	0.9	8.91	0.73	7.5	0.82	8.4
	8-14	11	10.45	8.81	1.64	15.7	8.27	0.54	6.2	2.18	20.9
	15-21	14	8.78	6.57	2.21	25.2	5.57	1.00	15.2	3.21	36.6
	22-28	6	7.33	3.00	4.33	59.1	2.50	0.50	16.7	4.83	65.9
	1-7	12	8.58	7.17	1.41	16.5	7.00	0.17	2.3	1.58	18.4
Methylbromophos	8-14	16	9.19	8.69	0.50	5.4	8.56	0.13	1.4	0.63	6.8
	15-21	12	8.92	8.42	0.50	5.6	7.92	0.50	5.9	1.00	11.2
	22-28	17	9.65	8.77	0.88	9.2	8.59	0.18	2.0	1.06	11.0
	29-35	12	9.58	9.25	0.33	3.5	9.08	0.17	1.8	0.50	5.2
	36-42	13	9.00	8.08	0.92	10.3	7.69	0.39	4.8	1.31	14.5
Ethylbromophos	43-49	14	8.71	7.71	1.00	11.5	7.57	0.14	1.9	1.14	13.1
	1-7	17	9.65	8.65	1.00	10.4	8.35	0.30	3.4	1.30	13.4
	8-14	11	10.00	9.64	0.36	3.6	9.28	0.36	3.8	0.72	7.2
	15-21	16	10.00	9.38	0.62	6.2	9.25	0.13	1.3	0.75	7.5
	22-28	14	10.43	9.79	0.64	6.2	9.43	0.36	3.6	1.00	9.6
	29-35	15	10.47	9.53	0.94	8.9	9.33	0.20	2.1	1.14	10.8
	36-42	15	10.20	9.07	1.13	11.1	8.67	0.40	4.4	1.53	15.0
	43-49	18	9.83	8.72	1.11	11.3	8.61	0.11	1.3	1.22	12.4

Other males were mated with untreated virgin females according to the classical procedure for the dominant lethal mutation assay in mouse (Bateman, 1977).

Seven-day mating intervals were used for a total of seven weeks. Pregnant females were killed 14 days after verification of the vaginal plugs.

Statistical analysis was carried out by the Kastenbaum and Bowman tables and the χ^2 test.

RESULTS AND DISCUSSION

In bone marrow metaphases (table 1) the percentages of chromosome aberrations observed in animals treated with the insecticides were below the control level. Mitomycin C induced a great number of lesions, especially in the males killed 24 h after the treatment. In spermatogonia (table 2), both methyl and ethylbromophos increased, though not significantly, the frequency of damaged cells. No chromosome exchange was observed. The total number of aberrations induced by mitomycin C exceeded 30%.

The number of live embryos per female was low, sometimes less than 8, in the females mated with males receiving methylbromophos (table 3). Nevertheless, the frequency of deciduomata and the total foetal mortality were not significantly higher than in the control group. The number of live embryos is likely to be correlated to the low number of corpora lutea per female.

The females mated with males injected with ethylbromophos did not show any increase in foetal lethality before and after implantation. On the other hand, mitomycin C induced both types of foetal mortality, increasing the recovery period.

Although conducted under drastic conditions at very high doses (1 g/kg), our experiments did not reveal cytogenetic effects in bone marrow cells and spermatogonia or genetic effects in the dominant lethal mutation assay. These results confirm the data concerning the mutagenic properties of methylbromophos in microorganisms and in *Drosophila*. This also parallels the lack of carcinogenic and teratogenic action of long term treatments in mammals (Vettorazzi, 1976).

ACKNOWLEDGMENTS

This research was performed under contract with the Belgian Ministry of Public Health.

REFERENCES

- Bateman A (1977) The dominant lethal assay in the male mouse.
In: Kilbey B, Legator M, Nichols W, Ramel C (eds) Hand-
book of mutagenicity test procedures. Elsevier Press,
Amsterdam, p 325
- Benes V, Sram R (1969) Mutagenic activity of some pesticides in
Drosophila melanogaster. *Industr Med* 38:50-53
- Evans E, Breckon G, Ford C ((1964) An air drying method for
meiotic preparations from mammalian testes. *Cytogenetics*
3:289-294
- Gilot-Delhalle J, Colizzi A, Moutschen J, Moutschen-Dahmen M
(1983) Mutagenicity of some organophosphorus compounds at
the ade 6 locus of Schizosaccharomyces pombe. *Mutation Res*
117:139-148
- Lippens R, Claeys M, Wildemaue C, Van Larebeke N (1983)
Mutagenicity studies on 10 pesticides on trichloroethane
and on diaminobenzidine. *Mutation Res* 113:277-278
- Moutschen J, Moutschen-Dahmen M, Degraeve N (1981) Metrifonate
and dichlorvos: Cytogenetic investigations. *Acta Phar-
macol Toxicol* 49, Suppl 5:29-39
- Vettorazzi G (1976) State of the art of the toxicological
evaluations carried out by the joint FAO/WHO meeting on
pesticide residues. II. Carbamate and organophosphorus
pesticides used in agriculture and public health. *Residue
Rev* 63:1-43
- Received August 25, 1983; accepted September 20, 1983